

Camptothecin and Podophyllotoxin Derivatives

Inhibitors of Topoisomerase I and II – Mechanisms of Action, Pharmacokinetics and Toxicity Profile

Jörg T Hartmann¹ and Hans-Peter Lipp²

- 1 Department of Oncology/Hematology/Immunology/Pneumology/Rheumatology, Eberhard Karls University Tübingen, UKT – Medical Center II, Tübingen, Germany
- 2 Department of Clinical Pharmacy, Eberhard-Karls-University Tübingen, Tübingen, Germany

Contents

Abstract	209
1. Camptothecins	211
1.1 Mechanism of Action	211
1.2 Topotecan	212
1.2.1 Clinical Pharmacokinetics	213
1.2.2 Tolerability Profile	213
1.2.3 Important Drug Interactions	214
1.2.4 Oral Formulation	214
1.3 Irinotecan	214
1.3.1 Clinical Pharmacokinetics	214
1.3.2 Tolerability Profile	217
1.3.3 Important Drug Interactions	219
1.4 Exatecan	219
1.4.1 Clinical Pharmacokinetics	219
1.4.2 Tolerability Profile	219
1.5 Other Camptothecin Derivatives	219
2. Podophyllotoxins	220
2.1 Mechanism of Action	220
2.2 Etoposide	220
2.2.1 Clinical Pharmacokinetics	220
2.2.2 Tolerability Profile	221
2.3 Teniposide	224
2.3.1 Clinical Pharmacokinetics	224
2.3.2 Tolerability Profile	224
3. Dual Inhibitors of Topoisomerase I and Topoisomerase II	225
4. Conclusion	225

Abstract

Camptothecins represent an established class of effective agents that selectively target topoisomerase I by trapping the catalytic intermediate of the topoisomerase I-DNA reaction, the cleavage complex. The water-soluble salt camptothecin-sodium – introduced in early trials in the 1960s – was highly toxic in animals, whereas the semisynthetic derivatives irinotecan and topotecan did not cause haemorrhagic cystitis because of their higher physicochemical stability and solubility at lower pH values. Myelosuppression, neutropenia and, to a lesser extent, thrombocytopenia are dose-limiting toxic effects of topotecan. In contrast

to the structurally-related topotecan, irinotecan is a prodrug which has to be converted to SN-38, its active form. SN-38 is inactivated by conjugation, thus patients with Gilbert's syndrome and other forms of genetic glucuronidation deficiency are at an increased risk of irinotecan-induced adverse effects, such as neutropenia and diarrhoea.

The cytotoxic mechanism of podophyllotoxin is the inhibition of topoisomerase II. Common adverse effects of etoposide include dose-limiting myelosuppression. Hypersensitivity reactions are more common with etoposide and teniposide than with etoposide phosphate because the formulations of the former contain sensitising solubilisers. Leukopenia and thrombocytopenia occur in 65% and 80%, respectively, of patients after administration of conventional doses of teniposide. Anorexia, vomiting and diarrhoea are generally of mild severity after administration of conventional doses of topoisomerase II inhibitors. Clinical pharmacokinetic studies have revealed substantial interindividual variabilities regarding the area under the concentration-time curve values and steady-state concentrations for all drugs reviewed in this article. Irinotecan, etoposide and teniposide are degraded via complex metabolic pathways. In contrast, topotecan primarily undergoes renal excretion. Regarding etoposide and teniposide, the extent of catechol formation over time during drug metabolism may be associated with a higher risk for secondary malignancies.

Inhibitors of topoisomerase I and II are commonly used anticancer drugs that are active cytotoxic agents in a variety of hematological cancers and solid tumours. A number of new compounds have been developed recently. The topoisomerases have functions in DNA-replication, chromosome condensation and chromosome segregation, and are highly conserved enzymes essential for the survival of eukaryotic cells. These drugs are part of several treatment regimens approved for the treatment of different cancers (e.g. ovarian, lung, colorectal and testicular carcinoma, non-Hodgkin's lymphoma and CNS tumours). Topoisomerase I and II inhibitors differ in their pharmacological properties, and pharmacokinetic and toxicity profiles. Clinical pharmacokinetic studies have revealed substantial variability regarding the area under the concentration-time curve (AUC) and steady state concentrations for the topoisomerase I and II inhibitors.^[1-5] Whereas irinotecan, etoposide and teniposide move through complex metabolic pathways during drug degradation, topotecan is eliminated via renal excretion without major degradation. In the case of irinotecan, plasma concentrations of the active metabolite SN-38 cannot predict the overall rate of drug-associated adverse effects, based on SN-38-associated activity

in the gut lumen.^[4] The camptothecin derivatives, topotecan and irinotecan, interact with the enzyme topoisomerase I, whereas the podophyllotoxin derivatives, etoposide and teniposide, target the enzyme topoisomerase II, resulting in different DNA disorders.^[1] Topoisomerase I modulates the topological structure of DNA by inducing transient DNA breaks. These single-strand breaks help to remove excessive positive and negative DNA supercoils, which arise during DNA replication and transcription. The interaction between the camptothecins and the enzyme results in the formation of a topoisomerase I DNA complex.^[2]

In contrast to topoisomerase II, cellular levels of topoisomerase I are relatively independent of the cell cycle phase in normal tissues. Thus, topoisomerase I activity is only slightly increased during conditions of cellular proliferation in cells and tissues. However, higher constitutive activities of this enzyme can be detected in several tumour tissues (e.g. adenocarcinoma of the colon and rectum) compared with healthy tissues.^[6]

This review summarises the drug development, pharmacological, pharmacokinetic, therapeutic and safety aspects of topoisomerase I and II inhibitors, in particular the camptothecins, irinotecan, exatecan

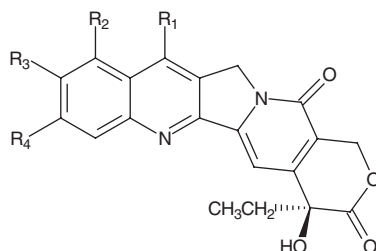
and topotecan, as well as the podophyllotoxins, etoposide and teniposide, and describes the possible supportive management and clinically relevant strategies to avoid toxicity by taking into account both clinical and pharmacological perspectives. Therefore, we searched the published literature on MEDLINE from January 1990 to December 2004, including only English-language articles. Searches were conducted using the keywords: 'human clinical trials', 'camptothecin', 'podophyllotoxin', 'topoisomerase I inhibitor', 'topoisomerase II inhibitor', 'pharmacology', 'pharmacokinetics', 'toxicity', 'side effects', 'irinotecan', 'topotecan', 'exatecan', 'teniposide', 'etoposide' and 'etoposide phosphate'. Additionally, case reports were selected for evaluation

and an extensive search of bibliographies from identified articles was performed. The literature was assessed for quality, methodology and outcome information.

1. Camptothecins

1.1 Mechanism of Action

Camptothecins (figure 1) were originally isolated from the wood, bark and fruit of the oriental tree, *Camptotheca acuminata* ('tree of joy'). Why the tree produces these highly toxic alkaloids is not known, but the most likely reason is that the toxins are part of a survival strategy to help combat herbivores. Among a large number of isolated plant con-



Compound	Molecular weight	R ₁	R ₂	R ₃	R ₄
Camptothecin	348.36	-H	-H	-H	-H
Topotecan	421.46 ^a	-H	-CH ₂ N(CH ₃) ₂	-OH	-H
Irinotecan	586.69 ^a	-CH ₂ CH ₃	-H	-O-C(=O)-N(CH ₂) ₄ -N(CH ₂) ₄ -	-H
SN-38	392.42	-CH ₂ CH ₃	-H	-OH	-H
9-Aminocamptothecin	363.38	-H	-NH ₂	-H	-H
9-Nitrocamptothecin	393.36	-H	-NO ₂	-H	-H
Lurtotecan (GI-147211)	518.57 ^a	-CH ₂ -N(CH ₂) ₄ -N(CH ₃)	-H	-OCH ₂ CH ₂ O-	-H
Exatecan	435.46 ^a	-CH(NH ₂)-CH ₂ -CH ₂ -	-H	-CH ₃	-F
Karenitecin	448.60	-H	-CH ₂ CH ₂ Si(CH ₃) ₃	-H	-H

^a Free base

Fig. 1. Chemical structures of camptothecins (reproduced from Garcia-Carbonero et al.,^[4] with permission).

Table 1. A comparison of topoisomerase I inhibitors: topotecan and irinotecan

Parameter	Topotecan	Irinotecan
Approved indication	Second-line treatment of advanced ovarian cancer and SCLC	Metastatic colorectal cancer
Dosage	1.5 mg/m ² IV d1–5, q3w Under investigation: 4 mg/m ² IV q1w (ovarian cancer)	350 mg/m ² IV, q3w 50–150 mg/m ² IV, q1w Under investigation: e.g. 200–250 mg/m ² q2w; 80–125 mg/m ² q1w
Clinical pharmacokinetics	Predominantly renal excretion with large amounts excreted as the parent drug	Prodrug that has to be converted to SN-38. Further metabolism includes SN-38 glucuronide, APC and NPC formation. Biliary excretion is greater than renal excretion.
	$t_{1/2}$ β : 2–3h (approximately 3h)	$t_{1/2\beta}$: approximately 14h

APC = 7-ethyl-10-[4-N-(5-aminopentanoic acid)-1-piperidino] carbonyloxycamptothecin; **IV** = intravenous; **NPC** = 7-ethyl-10-[(1-piperidino)-1-amino]-carbonyloxycamptothecin; **q_{xw}** = every 'x' weeks; **SCLC** = small-cell lung cancer; **$t_{1/2\beta}$** = terminal elimination half-life.

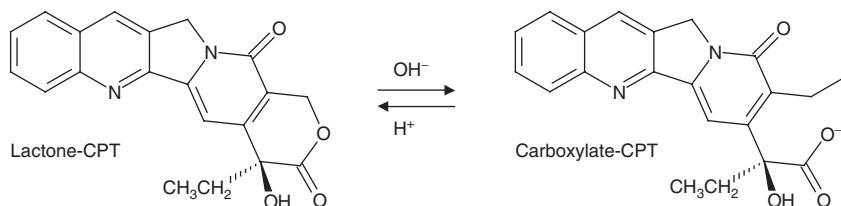
stituents, the naturally occurring alkaloid camptothecin (NSC94600) was identified as a highly potent inhibitor of topoisomerase I, which is overexpressed in many cancer cells. The water-soluble salt camptothecin-sodium, which was introduced in early preclinical trials in the 1960s, was highly toxic in animals.^[2] Hemorrhagic cystitis, leukopenia and thrombocytopenia were among the dose-limiting adverse effects of this agent. In addition, sterile haemorrhagic cystitis, myelosuppression, and gastrointestinal toxicity were common in patients during phase I studies.^[2] Further clinical trials with camptothecin-sodium were, therefore, discontinued in the 1970s.^[2] However, the semisynthetic derivatives irinotecan and topotecan (figure 1) are highly active in several malignancies and, because of their higher physicochemical stability and solubility at lower pH values, do not cause haemorrhagic cystitis.^[1] However, the drugs differ from each other in their approved therapeutic indications, recommended doses, toxicity profiles and clinical pharmacokinetics (table I).

Both irinotecan and topotecan contain a lactone moiety within their chemical structure (figure 2),

which can be hydrolysed non-enzymatically, resulting in the open-ring form.^[7] Under acidic conditions, the equilibrium between the biologically active lactone and the less active carboxylate form is generally shifted to the lactone form, whereas at physiological or higher pH values the lactone form becomes unstable because hydrolysis to the open form is favoured.^[7] In addition, the affinity of the carboxylate form for human serum albumin is estimated to be about 100-fold higher than that of the lactone form. As a consequence, >95% of the irinotecan dose is bound to serum albumin as a transiently inactive drug when irinotecan is given intravenously.^[8] Novel camptothecin derivatives, such as exatecan or lurtotecan (GI-147211), are more resistant to rapid hydrolysis due to their structural modifications (e.g. by the removal of the 20-OH group [figure 2]).^[4]

1.2 Topotecan

Topotecan, administered as an intravenous (IV) infusion of 1.5 mg/m² for 5 consecutive days of a 21-day cycle, has been approved for the second-line treatment of advanced ovarian and small-cell lung

**Fig. 2.** The carboxylate-lactone equilibrium of camptothecin (CPT) [reproduced from Loos et al.,^[9] with permission].

cancer (SCLC) in several countries.^[10] Preliminary data indicate that a dosage of 4 mg/m²/week may be associated with improved activity and reduced toxicity in patients with recurrent or persistent epithelial ovarian carcinoma.^[11]

1.2.1 Clinical Pharmacokinetics

After IV administration of conventional dosages (e.g. 1.5 mg/m²/day as a 30-minute infusion, on 5 consecutive days every 3 weeks), the half-life of topotecan is, on average, 3.2 hours (2.3–4.3 hours).^[10] The ratio of the AUC of the lactone to that of total topotecan appears to be relatively constant after IV administration, and averages about 0.20 over several hours, which means that only 20% of the total drug concentration in plasma represent the closed-ring active lactone form.^[10] The volume of distribution during steady-state conditions has been estimated to be 25–75 L/m², indicating extensive tissue binding.^[10,12] Red blood cells appear to act as a type of depot for topotecan (the lactone form), with steady-state concentrations that are almost 1.7-fold higher than those obtained in the plasma.^[13]

Following IV administration, topotecan is primarily excreted unchanged in the urine. Approximately 40% of the total dose (range 26–80%) can be recovered in the urine within 24 hours after the start of a 30-minute infusion, whereas up to 18% of dose can be recovered in the faeces. Urinary excretion of the carboxylate form is predominant when the urinary pH approximates a value of 6.9. Lower pH values may favour the lactone form. Despite these high urinary concentrations, topotecan itself does not cause urinary toxicity because of its high water solubility.^[10,12] Dose modification is warranted in patients with impaired renal function.^[14] Reduced dosages of 0.75 mg/m²/day and 0.5 mg/m²/day have been recommended in patients with reduced creatinine clearance (20–40 mL/min) who are either untreated or have been extensively pretreated, respectively.^[15] It has also been suggested that dosage adjustment may even be required if the creatinine clearance ranges from 40–60 mL/min. In those patients, the recommended starting dose should be 1.2 mg/m²/day IV on 5 consecutive days, in order to reduce the risk for severe myelosuppression.^[16] In general, topotecan should not be given to patients with severe renal insufficiency (e.g. creatinine clear-

ance <20 mL/min) until further data are available. In contrast, patients with impaired liver function and hyperbilirubinaemia do not appear to require topotecan dose modification. Preliminary study results indicate that topotecan is haemodialysable, when needed.^[17]

Hepatic metabolism of topotecan, mediated by cytochrome P450 (CYP) isoenzymes, is of minor quantitative importance. Metabolic pathways include N-dealkylation (resulting in N-demethyltopotecan) and glucuronidation.^[10]

The plasma protein binding of topotecan appears to be low (7–35%).^[10] In contrast to many other anticancer drugs, topotecan is able to pass through the blood-brain-barrier.^[18] More than 30% of the plasma concentration of topotecan can be recovered in the cerebrospinal fluid (CSF).^[18] Nevertheless, intrathecal drug administration (e.g. 0.4mg absolute dose) has been suggested to be advantageous when high concentrations in the CSF are warranted, in order to reduce the systemic toxicity. However, experience with this is thus far is based on preclinical study results and case reports.^[19,20]

1.2.2 Tolerability Profile

Myelosuppression, neutropenia and, to a lesser extent, thrombocytopenia are dose-limiting adverse effects of topotecan. Reversible non-cumulative neutropenia usually occurs between days 8 and 15 after a dosage of 1.5 mg/m² IV on 5 consecutive days. The nadir of the neutrophil count usually occurs on day 11, with recovery on day 21.^[21] Neutropenia, with cell counts <1.5 × 10⁹/L (grade 2) and <0.5 × 10⁹/L (grade 4), is observed in 70–97% of patients. In addition, 4–33% of patients treated with conventional doses of topotecan develop neutropenic fever during treatment cycles.^[21]

Thrombocytopenia, with platelet counts <50 × 10⁹/L (grade 3) and <25 × 10⁹/L (grade 4), occurs in 25–77% of patients, with a nadir on day 15 and recovery on day 21.^[21] Platelet transfusions are needed in 4–27% of patients.^[21] Anaemia, defined as a fall in haemoglobin levels to below 8 g/dL (grade 3) or 6.5 g/dL (grade 4), has been reported in 21–41% of patients during treatment cycles.^[21] As a consequence, red blood cell transfusions may be required in approximately 25% of treated patients.^[21] More extensive myelosuppression is ob-

Table II. Inhibitors and inducers of cytochrome P450 (CYP) 3A4-mediated biotransformation of topoisomerase I and II inhibitors

Interaction	Effect	Interacting agents
CYP3A4 inhibition	Increased topoisomerase inhibitor concentrations in the plasma	Amiodarone, cisapride, clarithromycin, erythromycin, ciclosporin, tacrolimus, delavirdine, ritonavir, fluconazole, voriconazole, itraconazole, ketoconazole, fluoxetine, grapefruit juice, nefazodone, terfenadine, verapamil, quinupristin/dalfopristin
CYP3A4 induction	Decreased topoisomerase inhibitor concentration in the plasma	Barbiturates, carbamazepine, oxcarbazepine, griseofulvin, nafcillin, phenytoin, primidone, rifabutin, rifampicin (rifampin), St John's wort, troglitazone

served in patients who have been extensively pre-treated with cytotoxic drugs. The extent of myelosuppression correlates significantly with both the total topotecan AUC and the topotecan lactone AUC.^[10] When granulocyte colony-stimulating factor (G-CSF) is given as prophylaxis against neutropenia, thrombocytopenia becomes the dose-limiting myelotoxic effect.^[21] In contrast to the structurally related irinotecan, gastrointestinal discomfort with topotecan is generally of mild severity.

1.2.3 Important Drug Interactions

Administration of cisplatin before topotecan has been shown to result in a sequence-dependent effect on the disposition of topotecan.^[22] Cisplatin-related acute changes in the glomerular filtration rate can temporarily alter topotecan clearance, with more severe myelosuppression as a consequence. Nevertheless, this sequence has sometimes been recommended in clinical trials, based on a potential exaggeration of the antineoplastic activity and the possibility of the use of G-CSF support to circumvent the adverse effect of neutropenia. However, severe thrombocytopenia may also occur, which limits the use of this treatment strategy.^[23-25] There is some evidence that potent inhibitors or inducers of CYP3A4 may be able to alter the clearance of topotecan; however, whether such interactions result in clinically relevant consequences has not yet been elucidated (table II).^[10]

1.2.4 Oral Formulation

In addition, an oral formulation of topotecan is undergoing clinical development.^[26-28] Administration schedules of oral topotecan 2.3 mg/m²/day for 5 days revealed similar efficacy results to topotecan 1.5 mg/m²/day IV for 5 days in patients with chemosensitive SCLC,^[26] whereas the IV schedule

was suggested to be favourable with regard to survival in patients with relapsed epithelial ovarian cancer.^[28] In general, oral topotecan appears as if it will be a convenient and well tolerated regimen for the treatment of defined solid tumours in the near future.^[26-28]

1.3 Irinotecan

Irinotecan has been approved for the first- and second-line treatment of advanced colorectal cancer, and has also been shown to be active in gastric cancer, non-small-cell lung cancer (NSCLC) and SCLC. Conventional dosages include 350 mg/m² IV every 3 weeks, 200–250 mg/m² IV every 2 weeks and 80–125 mg/m² IV weekly (see table I), whether given as a single agent or in combination (e.g. with 5-fluorouracil and folinic acid weekly). In contrast to the structurally related topotecan, irinotecan is a prodrug that needs bioactivation to SN-38 to become active.^[5,29,30]

1.3.1 Clinical Pharmacokinetics

Following IV administration, the bulky piperidino moiety at the C10 position of irinotecan is rapidly removed by carboxylesterases (CES). Of the three isoenzymes, CES2 plays the pivotal role in metabolism, followed by CES1A1 and CES3.^[31]

SN-38 is approximately 1000-fold more potent than its parent compound, irinotecan. There is a pH- and protein-dependent equilibrium between the active lactone and the inactive carboxylated forms for both irinotecan and SN-38.^[32]

SN-38 is inactivated by conjugation, which is primarily catalysed by the uridine diphosphate glucuronosyltransferase isoforms 1A7 (UGT1A7) and/or 1A9 (UGT1A9). The role of UGT1A1 has to be examined in more detail (figure 3).^[33,34] This

isoenzyme is also responsible for the glucuronidation of substrates such as bilirubin and valproic acid.^[35] Genetic enzyme deficiencies, such as Gilbert's syndrome or Crigler–Najjar syndrome type 1, result in impaired glucuronidation capacity. The molecular defects in Gilbert's syndrome are based on mutations within the five exons of the UGT1A gene locus.^[35] The incidence of Gilbert's syndrome ranges from 0.5 to 15% in different ethnic groups, which explains why there is significant variability in UGT1A activity in human livers; this variability can result in a 17-fold interindividual difference between the minimum and maximum rates of SN-38 glucuronidation.^[35] Patients with Gilbert's syndrome and other forms of genetic glucuronidation deficiency are at an increased risk of irinotecan-induced adverse effects, particularly gastrointestinal toxicity and leukopenia, if conventional doses are maintained.^[36] Genotype screening

is becoming increasingly feasible but is not yet routinely used. Thus, empirical irinotecan dose modification or the selection of another anticancer drug appears to be appropriate in patients with evidence of poor glucuronidation capacity (e.g. based on low levels of bilirubin glucuronide conjugates in the plasma).^[37,38] In addition, drug interactions may occur if constitutive SN-38 glucuronidation capacity is modified (e.g. by drugs such as phenytoin). However, the relative extent of such an interaction has not been clearly elucidated.^[39,40]

Irinotecan, SN-38 and its glucuronide metabolite, SN-38 glucuronide (SN-38G), are primarily excreted into the bile by the canalicular multispecific organic anion transporter (cMOAT, also known as multidrug resistance protein 2 [MRP2]), a member of the adenosine triphosphate group of transporters, but are also excreted by P-glycoprotein (also known as the multidrug resistance glycoprotein [MDR]).^[42]

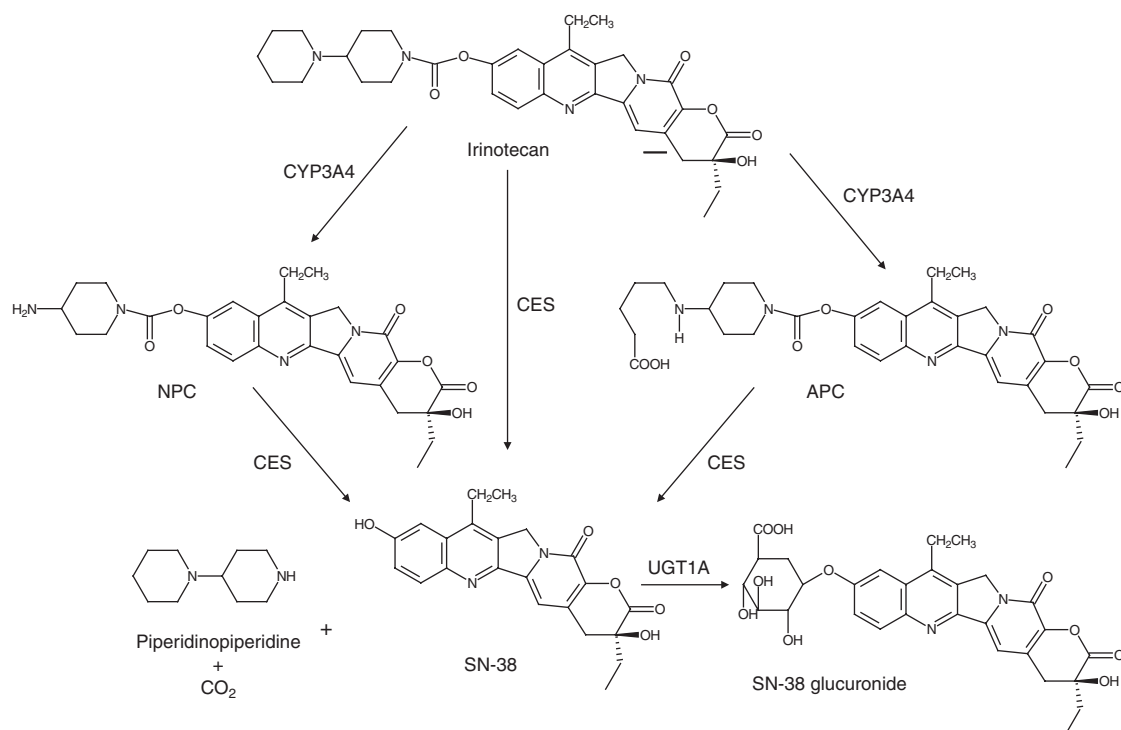


Fig. 3. Irinotecan is oxidised by the cytochrome P450 (CYP) 3A4 isoenzyme to produce 7-ethyl-10-[4-N-(5-aminopentanoic acid)-1-piperidino] carbonyloxycamptothecin (APC) and 7-ethyl-10-[4-(1-piperidino)-1-amino]-carbonyloxycamptothecin (NPC). NPC, APC and irinotecan are metabolised by carboxylesterases (CES) to produce the active metabolite, SN-38 (7-ethyl-10-hydroxycamptothecin). SN-38 is inactivated by uridine diphosphate glucuronosyltransferase isoform 1A1 (UGT1A) to SN-38 glucuronide (reproduced from Sanghani et al.,^[41] with permission).

Therefore, inhibitors of cMOAT and P-glycoprotein, such as ciclosporin, are able to reduce the clearance of irinotecan and SN-38. According to the study results published by Chester et al.,^[43] coadministration of IV irinotecan and oral ciclosporin resulted in a >2-fold increase in the AUC of irinotecan, SN-38 and SN-38G. As a consequence, the ratio of SN-38 to irinotecan was not altered, suggesting a limited effect of this interaction on drug metabolism. During this coadministration, the maximum tolerated irinotecan dosage was 100 mg/m² every 2 weeks. Interestingly, dose-limiting diarrhoea could not be observed in this trial, which may be related to the low SN-38 and SN-38G levels excreted into the gut lumen via the bile.^[43]

SN-38G excreted into the bile can be deconjugated by bacterial glucuronidases in the gut to active SN-38.^[44] SN-38 itself is able to undergo enterohepatic circulation that results in a second plasma peak of SN-38 after IV drug administration.^[45] However, SN-38 released within the gut lumen has been suggested to be an important cause of delayed intestinal toxicity; in animal experiments, constitutive bacterial β -glucuronidase activity correlated with irinotecan-induced gut damage. In contrast, the prophylactic use of oral antibacterials (e.g. aminoglycosides, bacitracin or quinolones) resulted in attenuation of intestinal effects.^[46]

Aminopentane-carboxylic acid (APC) is a further metabolite of irinotecan (figure 3); it is formed by oxidation of the terminal piperidine moiety, catalysed by CYP3A4. APC itself is not hydrolysed to SN-38 and is only a weak inhibitor of topoisomerase I.^[47-49] However, potent CYP3A4 inducers (e.g. St John's wort, carbamazepine and phenytoin) or inhibitors (e.g. itraconazole, ketoconazole) may alter the pharmacokinetics of irinotecan significantly.^[50] Other identified metabolites include 7-ethyl-10-(4-amino-1-piperidono)carbonyloxycamptothecin (NPC), 5-hydroxyirinotecan and RPR112526 (a decarboxylated product of the acid form of irinotecan lactone), which are altogether of minor quantitative importance.^[47-49,51] However, based on the fact that different enzymes, namely CES2, UGT1A and CYP3A4, contribute significantly to irinotecan metabolism, large interindividual variability may be observed regarding the AUC of SN-38; this variability has been estimated to be as high as 70% among

cancer patients treated with the same dose of irinotecan.^[52]

After IV infusion of irinotecan over 30–90 minutes the estimated half-life of the parent compound and SN-38 ranges from 5 to 14 hours. In contrast, continuous 5-day IV drug infusion schedules, which are not yet being commonly used, result in prolonged half-lives (approximately 27 and 30 hours for irinotecan and SN-38, respectively).^[47,48,53] Following IV administration, approximately 30% and 62% of the dose are excreted via the urine and faeces, respectively. Among excreted products, irinotecan accounts for 55% of the administered dose, followed by APC (10.5%), SN-38 (8.7%), SN-38G (3.3%) and NPC (1.5%).^[52] Based on the importance of hepatic and biliary clearance, a negative correlation between serum bilirubin concentrations and the total body clearance of irinotecan has been observed. Following one case report in a patient with moderately impaired liver function,^[54] it was calculated that, in order to achieve half-lives and maximum concentration values for irinotecan and SN-38 that were comparable to those observed in patients with normal liver function, the dose would have to be reduced to 100 mg/m² instead of the planned 350 mg/m² IV every 3 weeks. However, the corresponding AUCs were increased to a greater extent than expected, resulting in more severe leukopenia and delayed diarrhoea. The authors concluded that the dose should be further reduced to 30 mg/m² to improve tolerability in patients with moderate liver dysfunction.^[54] In addition, phase I and pharmacokinetic study results revealed that patients with elevated direct bilirubin levels (up to 7 mg/dL) had an increased frequency of dose-limiting adverse effects, even though the starting dose was 145 mg/m² IV instead of 350 mg/m² IV.^[55]

In conclusion, more detailed studies concerning irinotecan dose modification in patients with liver dysfunction are urgently needed. According to preliminary pharmacokinetic study results, standard IV doses of irinotecan every 3 weeks (350 mg/m²) can be used in patients with bilirubin levels up to 1.5-fold the upper limit of normal (ULN), whereas 200 mg/m² IV should not be exceeded in patients with bilirubin levels 1.5–3.0-fold the ULN.^[56,57]

The absolute bioavailability of irinotecan after oral administration is low and variable (10–20%).

Table III. Irinotecan-associated gastrointestinal discomfort

Characteristic	Acute (early) diarrhoea	Subacute (late) diarrhoea
Onset	On day of infusion	>24h after irinotecan infusion
Pathophysiology	Cholinergic syndrome	Hypersecretory in nature
Medical intervention	Prophylaxis: 0.25mg atropine Treatment: 0.25–1mg atropine	Prophylaxis (under debate): oral neomycin activated charcoal β-glucuronidase-inhibitors Treatment: loperamide (intensified) racecadodril octreotide opium tincture budesonide oral alkalinisation

Transintestinal transport of irinotecan and SN-38 by P-glycoprotein and CYP-mediated first-pass effects in the intestine account for this phenomenon.

1.3.2 Tolerability Profile

Myelosuppression and delayed diarrhoea represent the primary dose-limiting adverse effects of irinotecan.^[5,29,30] The dose-limiting adverse effects of irinotecan depend largely on the dose schedule. Myelosuppression, particularly leukopenia and neutropenia, and more rarely thrombocytopenia and anaemia, has been observed following conventional IV doses. Gastrointestinal toxicity, particularly diarrhoea, is also common and can be distinguished as acute or subacute forms.

Leukopenia

Leukopenia is a dose-limiting adverse effect of irinotecan. Weekly IV doses (e.g. 100–125 mg/m²) appear to produce a slightly greater incidence of grade 3–4 neutropenia compared with 3-weekly schedules (350 mg/m²; 16–28% vs 14–22%).^[4,32] The median leucocyte nadir occurs on day 15, with recovery 8 days later. Severe anaemia (haemoglobin concentrations <8 g/dL) and severe thrombocytopenia (platelet count <50 × 10⁹/L) occur in 15% and 2% of patients, respectively. In addition, some patients may develop eosinophilia during irinotecan treatment.^[48,55]

Diarrhoea

Besides leukopenia, diarrhoea is the most important dose-limiting adverse effect related to irinotecan treatment. Acute diarrhoea occurs very early after drug administration and appears to be due to

direct inhibition of the enzyme acetylcholinesterase.^[58] The delayed-onset form of diarrhoea starts some days after drug administration, is hypersecretory in nature and its severity correlates with the concentrations of the active compound SN-38 in the plasma and bowel.^[59]

The acute form of diarrhoea is short lasting and is part of the 'cholinergic syndrome' that is often accompanied by abdominal cramps, sweating, salivation, visual disturbances and lacrimation.^[32] The recommended dose of atropine is 0.25mg IV for prevention or 0.25–1.0mg for acute treatment of patients with early cholinergic symptoms^[32] (table III). As such symptoms have not been observed with other camptothecin derivatives, it can be speculated that these adverse effects are restricted to irinotecan, whose piperidino group bears some structural similarity to the potent nicotine receptor stimulant, dimethylphenylpiperazinium.^[58] Coadministration of irinotecan and oxaliplatin on the same day has been shown to increase the occurrence of cholinergic syndrome.^[60,61]

Delayed-onset diarrhoea of all grades of severity can be observed in nearly 90% of patients during the first three treatment cycles containing irinotecan.^[32] The 'cholera-like syndrome' usually begins several days after completion of the infusion.^[59] Grade 3–4 diarrhoea (grade 3: passing of 7–9 stools per day, incontinence or severe cramps; grade 4: passing of ≥10 stools per day, grossly bloody stools or a need for total parenteral nutrition) has been described in about 30–40% of patients, irrespective of whether they were treated on a weekly basis or every 3 weeks, with an average onset on day 6 (range: days

2–12).^[59] Of the four major pathophysiological mechanisms of diarrhoea (osmotic, secretory, altered motility and exudative), irinotecan-induced watery diarrhoea appears to be primarily secretory, based on abnormal ion transport in the intestinal epithelial cells.^[59,62]

There is increasing evidence that the extent and severity of irinotecan-induced gastrointestinal toxicity correlates with the concentration of the active compound SN-38 in the plasma and bowel.^[8,44] The role of irinotecan pharmacokinetics in the plasma with regards to the prediction of the severity of irinotecan-induced diarrhoea has been highlighted by the introduction of a 'biliary index', which is defined as the calculated product of the AUC ratio of SN-38 to SN-38G and the total AUC. A preliminary study suggested using preventive measures very early in the course of treatment when the biliary index exceeded a defined value (e.g. 3.5 hmg/L). However, this recommendation did not acquire broad acceptance because the biliary index cannot predict metabolic processes within the gut lumen.^[8,63]

After biliary excretion, SN-38G undergoes de-conjugation by bacteria-derived β -glucuronidase in the bowel. As a consequence, a strategy that has been proposed for reducing irinotecan-induced subacute diarrhoea is the inhibition of the intestinal microflora (e.g. with antibacterial agents or experimental selective β -glucuronidase inhibitors, such as saccharic acid 1,4-lactone).^[64] According to results of a pilot project, neomycin prophylaxis was associated with less severe forms of subacute diarrhoea in six of seven treated patients compared with controls.^[65] Besides bioactivation through bacterial β -glucuronidases, approximately 30% of the irinotecan dose is excreted via the bile in unchanged form, and thus, may be directly converted to SN-38 in the bowel by intestinal CES.^[66] In conclusion, nonabsorbable inhibitors of β -glucuronidase, as well as of intestinal CES, could be of value to effectively circumvent irinotecan-induced delayed diarrhoea.^[44]

Based on the fact that the equilibrium between the active lactone form and the ring-opened carboxylate form of irinotecan is pH dependent, oral alkalinisation with a mixture consisting of sodium bicarbonate (2.0 g/day), magnesium oxide (2.0–4.0 g/day), water (pH ≥ 7.2 , 1.5–2 L/day) and ursodeox-

ycholic acid (300 mg/day), combined with 'controlled' defecation was used in a phase II trial to reduce subacute gastrointestinal toxicity.^[67] The anticancer activity of irinotecan was maintained, whereas the incidence of delayed diarrhoea was significantly reduced compared with a non-randomised control group.^[67,68]

The efficacies of several symptomatic antidiarrhoeal drug treatment regimens, including intensified loperamide, octreotide, racecadodril and budesonide, have been extensively assessed (table III). Loperamide is recommended when the first signs of subacute, late-onset diarrhoea occur;^[59] loperamide not only delays small intestinal and whole gut transit, but also has some antisecretory activity in the human jejunum and colon.^[59] The starting dose is 4mg, followed by 2mg every 2 hours, which is continued until the diarrhoea has stopped for at least 12 hours. However, general premedication with loperamide is not recommended. If loperamide alone is not effective enough, racecadodril 100mg three times daily can be added to treatment. Racecadodril belongs to a group of drugs that block cyclic adenosine monophosphate-mediated hypersecretion in the gut by inhibiting the intestinal enzyme, enkephalinase.^[69] The somatostatin analogue octreotide is effective, particularly in loperamide-refractory patients.^[70] Subcutaneous dosages of 100–150 μ g three times daily with dose escalation up to 500 μ g every 8 hours have been used successfully for at least 48–96 hours and have resulted in an improvement of diarrhoea by at least one WHO toxicity grade. Other second-line agents include tincture of opium.^[71,72]

Oral budesonide has also been proposed to be beneficial in patients with subacute diarrhoea. Because of its 90% first-pass-mediated degradation in the liver, its systemic glucocorticoid activity is low.^[73] Budesonide controls symptoms of diarrhoea in most patients with inflammatory bowel disease.^[73] Preliminary data have suggested that the use of budesonide in patients with irinotecan-induced diarrhoea could reduce the severity of symptoms.^[74] In addition, in a phase III trial budesonide 3mg three times daily significantly prevented irinotecan-induced diarrhoea.^[75] As a consequence, budesonide appears to be an add-on option in patients who do not respond to high-dose oral loperamide.^[75,76] Very

recently, activated charcoal has been suggested to be particularly beneficial for the prevention of irinotecan-induced diarrhoea. Activated charcoal may be able to absorb free luminal SN-38, thereby reducing irinotecan-related intestinal toxicity. According to the results of a phase II trial, administration of activated charcoal resulted in a reduction in the occurrence of grade 3 to 4 diarrhoea, diminished the need for antidiarrhoeal medication consumption and allowed for increased irinotecan dose intensity.^[77] However, whether activated charcoal is more advantageous than other methods of diarrhoea prevention needs further investigation.

Oral immunomodulators, such as interleukin-15 or Kampo medicine, may also be helpful to reduce irinotecan-related diarrhoea; however, randomised clinical trials are needed to assess their efficacy compared with placebo.^[71]

1.3.3 Important Drug Interactions

St John's wort (300mg three times daily, starting 14 days before irinotecan administration) reduced the AUC of the active metabolite of irinotecan, SN-38, by 42% and diminished the severity of expected myelosuppression. Leukocyte and neutrophil counts were reduced by 8.6% and 4.3%, respectively, after coadministration of St John's wort with irinotecan, in contrast to reductions of 56% and 63%, respectively, following irinotecan monotherapy.^[50,78] Considering potential interactions between irinotecan and various enzyme-inducing antiepileptic drugs, such as phenytoin, carbamazepine, oxcarbazepine and phenobarbital, all have been shown to increase the median clearance of irinotecan lactone and significantly decrease the median systemic exposure to SN-38 lactone.^[50] The underlying mechanisms include CYP3A4 induction and UGT1A1 induction, as well as, in the case of phenobarbital, a possible increase in transporter activity leading to higher biliary drug excretion.^[39] As a consequence, non-enzyme-inducing antiepileptics (e.g. gabapentin and dexamethasone) appear to be preferable as comedication if clinically indicated in patients who receive irinotecan, in order to circumvent complex drug interactions.^[39,40,79]

In contrast, potent CYP3A4 inhibitors, such as ketoconazole and other triazoles, are able to reduce irinotecan clearance. However, the extent of dose

modification needed in the case of coadministration of irinotecan and CYP3A4 inhibitors such as triazole antimycotic agents and some macrolides has not yet been determined (table II).^[80]

1.4 Exatecan

Exatecan (DX-8951-f) is a novel synthetic water-soluble camptothecin derivative with a unique hexacyclic structure (figure 1). The recommended dosages of exatecan, according to phase II trials, are 0.5 mg/m²/day and 0.3 mg/m²/day as 30-minute infusions on 5 consecutive days for minimally pretreated and heavily pretreated patients, respectively (e.g. with anthracycline and taxane-refractory metastatic breast cancer, respectively).^[81]

In contrast to irinotecan, exatecan does not require metabolic activation. According to *in vitro* experiments in various cancer cell lines, exatecan may be 6- and 28-fold more active as an anticancer drug than SN-38 and topotecan, respectively.^[82] In addition, preclinical studies have revealed that the therapeutic index (maximum tolerated dose : minimum effective dose) of exatecan is approximately 2- to 10-fold greater than the corresponding values for irinotecan and topotecan.^[83] In addition, exatecan may even be active in the presence of P-glycoprotein-mediated multidrug resistance.^[4]

1.4.1 Clinical Pharmacokinetics

Pharmacokinetic studies indicate that, following a 30-minute IV exatecan infusion, the elimination half-life may be, on average, 8 hours. The amount of unchanged exatecan recovered in urine within 24 hours ranged from 1% to 16%, as the hepatic metabolism of this drug is extensive. This is in contrast to the structurally related topotecan.^[81,84,85]

1.4.2 Tolerability Profile

Thus far, the dose-limiting adverse effects of exatecan have been identified as being neutropenia and liver dysfunction, whereas gastrointestinal discomfort is a rare event.^[81,84,85]

1.5 Other Camptothecin Derivatives

Several other camptothecin analogues are currently under investigation, such as the water-soluble derivative lurtotecan (GI-147211) and the poorly water-soluble analogues IDEC-132 (9-amino-

camptothecin) and rubitecan (9-nitrocamptothecin), which can be given orally.^[86-89] Further trials with topoisomerase I inhibitors include the feasibility of oral administration of topotecan and irinotecan, the use of a liposomal lurtotecan formulation (OSI 211 [NX211]) and the use of a pegylated derivative or other forms of the naturally occurring camptothecin, which are more soluble in aqueous solutions even at low pH values and may allow dose intensification.^[90,91]

2. Podophyllotoxins

2.1 Mechanism of Action

Etoposide and teniposide (table IV) represent semisynthetic derivatives of podophyllotoxin, which was originally isolated from the root of the Indian Podophyllum plant. After extensive isolation procedures the most effective 'antineoplastic' factor was identified as being 4'-demethylepipodophyllin benzyldene glucoside (DEPBG). Etoposide, its water-soluble derivative etoposide phosphate, and teniposide, are semisynthetic analogues of DEPBG that have increased antineoplastic activity (figure 4). Etoposide is active in testicular carcinoma, non-Hodgkin's lymphoma and other lymphomas, ovarian carcinoma, SCLC and NSCLC, and neoplasms of unknown origin.^[92] The pivotal indications for teniposide include malignant lymphoma and glioblastoma.^[93,94]

Compared with the anthracyclines or rebeccamycin analogues, which also act as topoisomerase II inhibitors, etoposide and teniposide are neither able to intercalate sterically within DNA strands nor is their use associated with the chronic cardiotoxicity seen with anthracyclines.^[96] Both drugs appear to act directly on the enzyme, which results in the stabilisation of a transient covalent complex formed between the enzyme and the DNA at break sites. During the presence of the epipodophyllotoxin derivatives, the enzyme-DNA intermediate cannot be reversed, resulting in DNA double strand breaks. Several studies have suggested that the activity of etoposide may be schedule dependent. The antiproliferative effect of etoposide on tumour cells appears to be greater when it is administered over several consecutive days rather than on a single day.

At higher doses, podophyllotoxins may also act as spindle poisons.^[97-99]

2.2 Etoposide

2.2.1 Clinical Pharmacokinetics

After IV administration of etoposide (e.g. 150 mg/m²) the peak plasma concentration and half-life average 20 µg/mL and 7.1 hours, respectively. The drug clearance and volume of distribution have been determined to be 16 mL/min/m² and 7.1 L/m², respectively. Regarding plasma concentrations of etoposide, IV etoposide phosphate and IV etoposide have been shown to be bioequivalent, irrespective of whether conventional or intensified dose schedules have been used.^[100,101] After IV administration, the prodrug etoposide phosphate undergoes rapid and complete hydrolysis catalysed by alkaline phosphatase. This conversion is not saturable even at high IV doses of up to 1200 mg/m² infused over 2 hours.^[102,103]

If etoposide is taken orally, the absolute bioavailability is dependent on the dose. When the total etoposide doses are 50–100mg and 400mg, the average values have been determined to be about 75% and 50%, respectively.^[104-107] If etoposide phosphate is taken orally, the average values appear to be somewhat higher.^[108] However, the advantage of oral etoposide phosphate may be diminished when the prodrug is administered together with acid beverages, rather than with concomitant acid suppressive medication, based upon the importance of intestinal pH-dependent alkaline phosphatase activity during bioactivation.^[109] Considering oral treatment with etoposide, marked interindividual variability has been observed in patients (e.g. children with relapsed acute lymphoblastic leukaemia).^[110] As a consequence, therapeutic drug monitoring has been suggested to be beneficial in order to maintain effective cytotoxic drug concentrations over time and to simultaneously reduce the risk for systemic toxicity.^[111] However, the translation of scientific data into everyday clinical practice remains a great challenge.

Approximately 96% of the dose of etoposide is bound to plasma proteins, whereas the unbound fraction averages 4%. There is a higher risk of myelotoxicity when the unbound fraction is in-

creased by factors such as hyperbilirubinaemia or hypoalbuminaemia, which may be of clinical relevance in patients with hepatic dysfunction or cachexia-inducing tumours.^[112-118] In addition, the interindividual variability of drug concentrations has been suggested to be of importance in patients who receive high-dose IV etoposide followed by reinfusion of autologous peripheral blood stem cells.^[119] If high drug concentrations persist over a longer period of time, the success of engraftment may be severely impaired. As a consequence, plasma drug concentration monitoring might be useful in order to identify patients at higher risk of graft failure.^[119,120]

The renal clearance of etoposide is about 30–40% of the total plasma clearance. Hepatic etoposide metabolism is primarily mediated by CYP3A4, and less so by CYP3A5, and results in the formation of a catechol metabolite (3-hydroxyetoposide) via o-demethylation, which is able to undergo sequential one-electron oxidation reactions resulting in the corresponding semiquinone and quinone moieties (figure 5). In this context, it is still a matter of debate as to what extent catechol formation may contribute to the late adverse effects of etoposide, particularly its leukaemogenic activity.^[121,122] Further metabolic pathways of minor quantitative importance include glucuronidation and hydroxyacid formation. Dose reductions of 33% and 50% have been recommended for patients with creatinine clearance values of 15–25 mL/min and <15 mL/min, respectively.^[113,123] In patients with obstructive jaundice and a reduced glomerular filtration rate, a 50% empiric dose reduction may be required.^[124]

So far, haemodialysis appears to be only moderately effective for the elimination of etoposide from systemic circulation.^[125]

In patients with brain metastases, high IV doses of etoposide may be needed in order to achieve adequate drug concentrations in the CSF.^[126] Intrathecal drug administration may be an alternative able to reduce the systemic toxicity associated with dose intensive chemotherapy; however, clinical experience is so far very scarce and is only based on case reports.^[127]

A large interindividual variability in etoposide plasma concentrations has been observed following conventional doses.^[128] As a consequence, some authors have suggested that by maintaining defined plasma concentrations of etoposide, treatment success might be optimised. For example, in patients with SCLC receiving etoposide 500 mg/m² IV as an infusion over 24 hours or as a 5-day schedule with 100 mg/m²/day IV, the latter was associated with a higher response rate regarding partial remission as well as a favourable increase in median survival time, which might be correlated with longer exposure to concentrations exceeding 1 µg/mL.^[128,129] However, more extended etoposide concentrations over time are also associated with a higher myelosuppressive burden, which limits, for example, either prolonged drug infusions or oral etoposide administration in routine practice.^[111,113]

2.2.2 Tolerability Profile

A common adverse effect of etoposide and etoposide phosphate is dose-limiting myelosuppression. Leukopenia is the most common adverse effect

Table IV. Comparison of topoisomerase II inhibitors: etoposide and teniposide

Parameter	Etoposide (and etoposide phosphate)	Teniposide
Approved use	SCLC, NSCLC Lymphoma, AML Choriocarcinoma, advanced ovarian carcinoma Testicular cancers	CNS tumours, malignant lymphoma, bladder cancer
Conventional dosage recommendations	50–100 mg/m ² , d1–5 IV 100–200 mg/m ² /d PO, d1–5	30 mg/m ² , d1–5 40–50 mg/m ² , d1–3 100–130 mg/m ² , q1w
Pharmacokinetics	t _{1/2β} : 6–8h Metabolism via CYP3A4 Plasma protein binding: >94%	t _{1/2β} : 8h Metabolism via CYP3A4 Plasma protein binding: >99%

AML = acute myelogenous leukaemia; **CYP** = cytochrome P450; **IV** = intravenous; **NHL** = non-Hodgkin's lymphoma; **NSCLC** = non-small-cell lung cancer; **PO** = oral; **q1w** = every week; **SCLC** = small-cell lung cancer; **t_{1/2β}** = terminal elimination half-life.

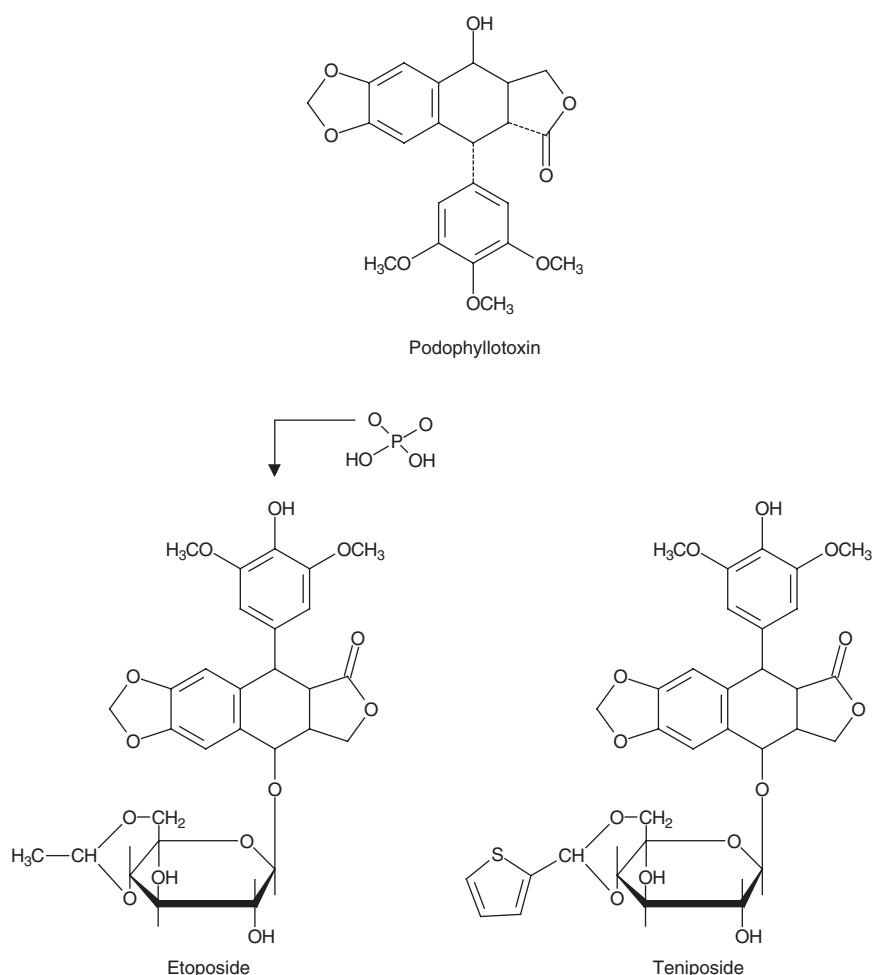


Fig. 4. The topoisomerase II inhibitor etoposide, its water-soluble derivative etoposide phosphate, and teniposide are semisynthetic derivatives of the natural compound podophyllotoxin.^[95]

associated with oral and IV etoposide. Nadirs in neutrophil counts generally occur within 7–14 days after administration.^[130] Thrombocytopenia occurs in 23% of etoposide-treated patients and about 9% develop platelet counts of $<50 \times 10^9/\text{L}$. Anorexia, vomiting and diarrhoea are generally of mild severity after administration of conventional doses of etoposide.^[131] Mucositis is more severe in patients who receive IV doses of etoposide up to 1000 mg/m^2 than those who receive lower dosages.^[120]

Hypersensitivity reactions are more common with etoposide and teniposide than with etoposide phosphate because the formulations of the former

agents contain sensitising solubilisers.^[132,133] The characteristic features of hypersensitivity reactions that occur after IV etoposide administration include bronchospasms, facial flushing, rashes, dyspnoea, fever, chills, tachycardia, chest tightness, cyanosis and changes in blood pressure (hypotension and hypertension).^[134,135] However, very severe forms of hypersensitivity reaction, such as Stevens-Johnson syndrome, are extremely rare.^[136] Anaphylactic-like reactions have occurred in 0.7–2% of patients after etoposide administration. With very few exceptions, patients recover quickly when the drug infusion is stopped immediately.^[137] Hypersensitivity reactions

to etoposide or teniposide appear to be type I hypersensitivity (IgE mediated), with an onset of symptoms that usually occurs within minutes after IV administration. Several reports have suggested that premedication with an antihistamine and/or a corticosteroid may prevent further hypersensitivity reactions, even in patients with a history of previous complications.^[138] However, this strategy should not be considered as routine practice when patients have previously had a severe hypersensitivity reaction, such as long-lasting bronchospasms or severe hypotension. Based on several case reports, etoposide could successfully be restarted in 78% of patients who had had experienced a hypersensitivity reaction, by slowly infusing the drug after premedication with an antihistamine and a corticosteroid.^[139]

Hypersensitivity reactions to etoposide and teniposide are primarily related to the adjuvants used in

the parenteral formulations, rather than to the drugs themselves. Etoposide formulations for parenteral use contain several adjuvants, including polysorbate 80, benzyl alcohol and polyethylene glycol, because the drug is rather insoluble in aqueous solutions. Polysorbate 80 has particularly been implicated as a causative agent for hypotension and metabolic acidosis, especially at higher doses.^[132] In contrast, the structurally-related etoposide phosphate is highly soluble in aqueous solutions and, thus, no solubilising adjuvants are needed. Preliminary data suggest that the incidence of hypersensitivity reactions is lower with etoposide phosphate than with etoposide, which strengthens the hypothesis that adjuvants play a major role in the development of allergic reactions. According to case reports, patients who had a type I hypersensitivity reaction to etoposide could be successfully retreated with etoposide phosphate.^[139,140]

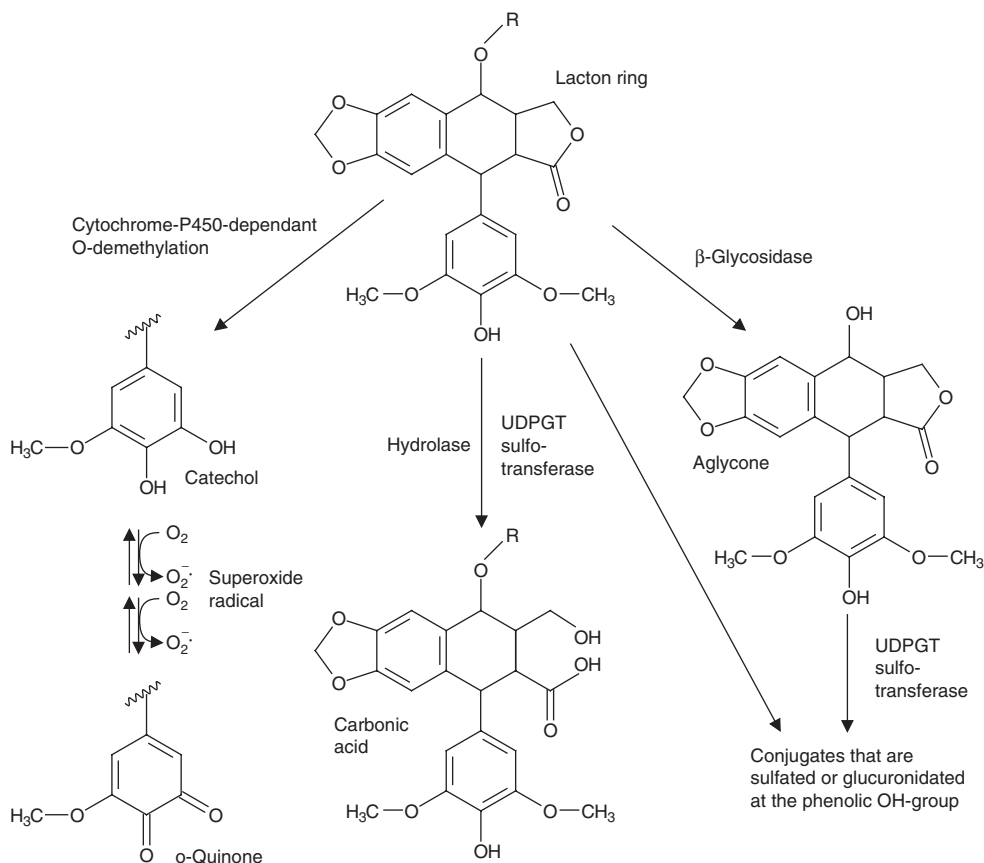


Fig. 5. Enzymatic biotransformation of the podophyllotoxin derivatives. **UDPGT** = uridine diphosphate-glucuronosyltransferase.^[95]

However, cross reactivity to etoposide has been observed in patients with hypersensitivity reactions to teniposide, suggesting that the allergic reactions are not exclusively restricted to the use of the solvents. In addition, hypersensitivity reactions have also been reported after the use of oral etoposide.^[141]

Neurotoxic symptoms are occasionally observed following treatment with etoposide (and teniposide), particularly at higher doses. Adverse nervous system effects, including headache, transient mental confusion and vertigo, may be related to the blood alcohol concentration, since teniposide and etoposide formulations both contain this solvent.^[142] Peripheral neuropathy, mainly mild and infrequent, has been described after conventional doses of etoposide and teniposide. However, more severe forms of peripheral neuropathy have been reported during coadministration of etoposide and vinca alkaloids.^[143,144]

Based on animal experiments, etoposide and teniposide can be classified as potential carcinogens.^[145,146] In addition, both drugs appear to be important risk factors for the development of secondary acute myelogenous leukaemia, as they have considerable leukaemogenic activity.^[147-156] Of 119 patients with advanced NSCLC, 24 survived for >1 year after treatment with etoposide and cisplatin, with or without vindesine. Of these 24 patients, four developed secondary acute myelogenous leukaemia at 13, 19, 28 and 35 months after the start of treatment. All four patients received a two-fold greater cumulative dose of etoposide (6.8 vs 3.0 g/m²) than the rest of the study cohort.^[132] Characteristically, podophyllotoxin-related secondary acute myelogenous leukaemia develops after a rather short latency (2–3 years) and differs from the malignancies caused by other drugs (e.g. alkylating agents) by its unique molecular marker, a balanced translocation involving the mixed-lineage leukaemia (MLL) gene on chromosome 11 ('11q23 abnormalities').^[157] Southern blot analysis of enzyme-digested DNA from etoposide-treated cell lines and from peripheral blood cells after treatment with etoposide showed frequent rearrangements of MLL, but not of other genes. Altogether, the etoposide-related incidence of secondary acute myelogenous leukaemia in three retrospective case series has been described as ranging from 0.4% to 8.1%. Secondary leukaemias de-

veloped 9–68 months after the diagnosis of the first cancer.^[158]

In conclusion, podophyllotoxin-containing regimens carry a small but significant risk of secondary acute myelogenous leukaemia, which, *per se*, is difficult to treat.^[159] The risk may be increased by higher total cumulative doses (e.g. etoposide >2 g/m²), weekly or twice weekly schedules, the concomitant administration of drugs that inhibit DNA repair, concomitant radiotherapy or the use of high doses of cisplatin. It has, therefore, been recommended that higher cumulative doses of etoposide should be used cautiously in low-risk diseases.^[158]

2.3 Teniposide

2.3.1 Clinical Pharmacokinetics

After IV administration, teniposide is more extensively metabolised than etoposide. Plasma concentration monitoring has been proposed to be beneficial in patients receiving teniposide. For example, in one study the maintenance of steady-state teniposide concentration >12 µg/mL appeared to be important for higher clinical response rates in patients with recurrent leukaemia, lymphoma or neuroblastoma.^[160] In those patients whose steady-state concentrations were maintained at >12 µg/mL, there was a shrinkage of the tumour, whereas patients with lower steady-state concentrations had a poorer response. Similar to etoposide, teniposide is a substrate of CYP3A4.^[122] As a consequence, its clearance is increased by inducers of this enzyme, such as carbamazepine, phenobarbital, phenytoin, rifampicin (rifampin) and St John's wort (table II). In contrast, competitive inhibitors of this enzyme may decrease the clearance of the podophyllotoxin derivatives (table II).^[161,162]

The bioavailability of teniposide after oral administration is about 42% (range 20–71%). Although teniposide 50mg capsules have been suggested to be useful in clinical practice, no oral formulation has been approved thus far.^[163]

2.3.2 Tolerability Profile

Leukopenia and thrombocytopenia occur in 65% and 80%, respectively, of patients after administration of conventional doses of teniposide.^[94] Anorexia, vomiting and diarrhoea are generally of mild severity after administration of conventional doses

of teniposide.^[164] Some data suggest that the overall incidence of hypersensitivity reactions to teniposide may be as high as 50%, if all forms of hypersensitivity reaction are taken into account. In the case of teniposide, the solubilising adjuvant polyoxyl-35 castor oil has been implicated as the pivotal causative agent.^[132] However, in nine children who developed facial oedema and flushing after receiving teniposide, the drug itself degranulated basophils *in vitro* and caused histamine release, while polyoxyl-35 castor oil did not.^[132] Teniposide is about 10-fold more potent than etoposide in causing DNA damage *in vitro* and *in vivo*. In 21 of 733 children with acute lymphoblastic leukaemia who were in remission and who received maintenance therapy with teniposide once or twice weekly in combination with other anticancer drugs, the risk of secondary acute myelogenous leukaemia was about 12-fold higher than in patients who had been treated with less intensive schedules (e.g. a short course of teniposide for induction chemotherapy).^[155]

3. Dual Inhibitors of Topoisomerase I and Topoisomerase II

According to *in vitro* results, the administration of a topoisomerase I inhibitor in cancer cell lines resulted in a compensatory increase of topoisomerase II levels and *vice versa*.^[165] As a consequence, schedule-dependent use of topoisomerase I and II inhibitors has been suggested to improve efficacy. However, based on overlapping myelosuppressive adverse effects, such a combined use of, for example, topotecan and etoposide, has not yet been established in clinical practice.^[166-168] Based on preclinical study results, some drugs, such as F-11782, TAS-103, intoplicine and XR-5000, were considered encouraging dual topoisomerase I and II inhibitors; however, thus far none of these drugs have entered phase II or III trials.^[169-171]

4. Conclusion

Camptothecin and podophyllotoxin derivatives are commonly used anticancer drugs in clinical oncology. Clinical pharmacokinetic studies have revealed substantial interindividual variabilities regarding AUC values and steady-state concentrations for all of the reviewed drugs (irinotecan, topotecan,

etoposide and teniposide), which are – with the exception of topotecan – primarily because of the complex metabolic pathways involved in drug degradation. Some of the resultant metabolites contribute significantly to the observed toxicity of these drugs, which is of clinical concern. However, plasma concentrations of SN-38, the active metabolite of irinotecan, cannot predict gastrointestinal adverse effects accurately, because relevant levels of the active metabolite can also be formed in the gut by bacterial β -glucuronidases and intestinal CES. Regarding etoposide and teniposide, the extent of catechol formation over time during drug metabolism – although low – may be associated with a higher risk for secondary malignancies. In contrast, topotecan undergoes primarily renal excretion, with the parent drug being the most prominent substance recovered in the 24-hour urine. However, creatinine clearance varies considerably among patients, indicating that more precise renal function assessments than serum creatinine values are needed in order to accurately translate published recommendations for dose modification of topotecan into clinical practice. Medical oncologists have to be familiar with the clinical pharmacokinetics of topoisomerase I and II inhibitors and their possible drug interactions in order to modify the dose in time to avoid substantial subacute and late adverse effects.

Acknowledgements

We acknowledge the excellent assistance of Mrs Gabi Jany in the preparation of the manuscript.

No sources of funding were used to assist in the preparation of this review. The authors have no conflicts of interest that are directly relevant to the content of this review.

References

1. Malonne H, Atassi G. DNA topoisomerase targeting drugs: mechanisms of action and perspectives. *Anticancer Drugs* 1997; 8: 811-22
2. Potmesil M. Camptothecins: from bench research to hospital wards. *Cancer Res* 1994; 54: 1431-9
3. Iyer L, Ratain MJ. Clinical pharmacology of camptothecins. *Cancer Chemother Pharmacol* 1998; 42 Suppl.: S31-43
4. Garcia-Carbonero R, Supko JG. Current perspectives on the clinical experience, pharmacology, and continued development of the camptothecins. *Clin Cancer Res* 2002; 8: 641-61
5. Rothenberg ML. Topoisomerase I inhibitors: review and update. *Ann Oncol* 1997; 8: 837-55
6. Husain I, Mohler JL, Seigler HF, et al. Elevation of topoisomerase I messenger RNA, protein, and catalytic activity in human tumors: demonstration of tumor-type specificity and implications for cancer chemotherapy. *Cancer Res* 1994; 54: 539-46

7. Burris HA, Rothenberg ML, Kuhn JG, et al. Clinical trials with the topoisomerase I inhibitors. *Semin Oncol* 1992; 19: 663-9
8. Gupta E, Mick R, Ramirez J, et al. Pharmacokinetic and pharmacodynamic evaluation of the topoisomerase inhibitor irinotecan in cancer patients. *J Clin Oncol* 1997; 15: 1502-10
9. Loos WJ, Gelderblom HJ, Verweij J, et al. Gender-dependent pharmacokinetics of topotecan in adult patients. *Anticancer Drugs* 2000; 11: 673-80
10. Herben VM, ten Bokkel Huinink WW, Beijnen JH. Clinical pharmacokinetics of topotecan. *Clin Pharmacokinet* 1996; 31: 85-102
11. Levy T, Inbar M, Menczer J, et al. Phase II study of weekly topotecan in patients with recurrent or persistent epithelial ovarian cancer. *Gynecol Oncol* 2004; 95: 686-90
12. Herben VM, Schoemaker E, Rosing H, et al. Urinary and fecal excretion of topotecan in patients with malignant solid tumours. *Cancer Chemother Pharmacol* 2002; 50: 59-64
13. Loos WJ, Gelderblom H, Verweij J, et al. Red blood cells: a neglected compartment in topotecan pharmacokinetic analysis. *Anticancer Drugs* 2003; 14: 227-32
14. O'Reilly S. Topotecan: what dose, what schedule, what route? *Clin Cancer Res* 1999; 5: 3-5
15. O'Reilly S, Rowinsky EK, Slichenmyer W, et al. Phase I and pharmacologic study of topotecan in patients with impaired renal function. *J Clin Oncol* 1996; 14: 3062-73
16. Montazeri A, Culine S, Laguerre B, et al. Individual adaptive dosing of topotecan in ovarian cancer. *Clin Cancer Res* 2002; 8: 394-9
17. Herrington JD, Figueroa JA, Kirstein MN, et al. Effect of hemodialysis on topotecan disposition in a patient with severe renal dysfunction. *Cancer Chemother Pharmacol* 2001; 47: 89-93
18. Dennis MJ, Beijnen JH, Grochow LB, et al. An overview of the clinical pharmacology of topotecan. *Semin Oncol* 1997; 24: S5-18
19. Blaney SM, Heideman R, Berg S, et al. Phase I clinical trial of intrathecal topotecan in patients with neoplastic meningitis. *J Clin Oncol* 2003; 21: 143-7
20. Blaney SM, Cole DE, Godwin K, et al. Intrathecal administration of topotecan in nonhuman primates. *Cancer Chemother Pharmacol* 1995; 36: 121-4
21. Kollmannsberger C, Mross K, Jakob A, et al. Topotecan: a novel topoisomerase I inhibitor. *Pharmacology and clinical experience. Oncology* 1999; 56: 1-12
22. Rowinsky EK, Kaufmann SH, Baker SD, et al. Sequences of topotecan and cisplatin: phase I, pharmacologic, and in vitro studies to examine sequence dependence. *J Clin Oncol* 1996; 14: 3074-84
23. Breidenbach M, Rein DT, Schondorf T, et al. Hematological side-effect profiles of individualized chemotherapy regimen for recurrent ovarian cancer. *Anticancer Drugs* 2003; 14: 341-6
24. Saltz L, Sirott M, Young C, et al. Phase I clinical and pharmacology study of topotecan given daily for 5 consecutive days to patients with advanced solid tumors, with attempt at dose intensification using recombinant granulocyte colony-stimulating factor. *J Natl Cancer Inst* 1993; 85: 1499-507
25. Rowinsky EK, Grochow LB, Sartorius SE, et al. Phase I and pharmacologic study of high doses of the topoisomerase I inhibitor topotecan with granulocyte colony-stimulating factor in patients with solid tumors. *J Clin Oncol* 1996; 14: 1224-35
26. von Pawel J, Gatzemeier U, Pujol JL, et al. Phase II comparator study of oral versus intravenous topotecan in patients with chemosensitive small-cell lung cancer. *J Clin Oncol* 2001; 19: 1743-9
27. Creemers GJ, Gerrits CJ, Eckardt JR, et al. Phase I and pharmacologic study of oral topotecan administered twice daily for 21 days to adult patients with solid tumors. *J Clin Oncol* 1997; 15: 1087-93
28. Gore M, Oza A, Rustin G, et al. A randomised trial of oral versus intravenous topotecan in patients with relapsed epithelial ovarian cancer. *Eur J Cancer* 2002; 38: 57-63
29. Saigi E, Salut A, Campos JM, et al. Phase II study of irinotecan (CPT-11) administered every 2 weeks as treatment for patients with colorectal cancer resistant to previous treatment with 5-fluorouracil-based therapies: comparison of two different dose schedules (250 and 200 mg/m²) according to toxicity prognostic factors. *Anticancer Drugs* 2004; 15: 835-41
30. Hofheinz R, Hartung G, Samel S, et al. Adding weekly irinotecan to high-dose 5-fluorouracil and folinic acid (HD-5-FU/FA) after failure for first-line HD-5-FU/FA in advanced colorectal cancer: a phase II study. *Anticancer Drugs* 2002; 13: 999-1004
31. Sanghani SP, Quinney SK, Fredenburg TB, et al. Hydrolysis of irinotecan and its oxidative metabolites, 7-ethyl-10-[4-N-(5-aminopentanoic acid)-1-piperidino] carbonyloxy-camptothecin and 7-ethyl-10-[4-(1-piperidino)-1-amino]-carbonyloxy-camptothecin, by human carboxylesterases CES1A1, CES2, and a newly expressed carboxylesterase isoenzyme, CES3. *Drug Metab Dispos* 2004; 32: 505-11
32. Cersosimo RJ. Irinotecan: a new antineoplastic agent for the management of colorectal cancer. *Ann Pharmacother* 1998; 32: 1324-33
33. Carlini LE, Meropol NJ, Bever J, et al. UGT1A7 and UGT1A9 polymorphisms predict response and toxicity in colorectal cancer patients treated with capecitabine/irinotecan. *Clin Cancer Res* 2005; 11: 1226-36
34. Iyer L, King CD, Whittington PF, et al. Genetic predisposition to the metabolism of irinotecan (CPT-11): role of uridine diphosphate glucuronosyltransferase isoform 1A1 in the glucuronidation of its active metabolite (SN-38) in human liver microsomes. *J Clin Invest* 1998; 101: 847-54
35. Kraemer D, Scheurlen M. Gilbert disease and type I and II Crigler-Najjar syndrome due to mutations in the same UGT1A1 gene locus [in German]. *Med Klin (Munich)* 2002; 97: 528-32
36. Innocenti F, Undevia SD, Iyer L, et al. UGT1A1*28 polymorphism is a predictor of neutropenia in irinotecan chemotherapy [abstract 495]. *Proc Am Soc Clin Oncol* 2003; 22: 124
37. Ando Y, Saka H, Asai G, et al. UGT1A1 genotypes and glucuronidation of SN-38, the active metabolite of irinotecan. *Ann Oncol* 1998; 9: 845-7
38. Innocenti F, Iyer L, Ratain MJ. Pharmacogenetics of anticancer agents: lessons from amonafide and irinotecan. *Drug Metab Dispos* 2001; 29: 596-600
39. Crews KR, Stewart CF, Jones-Wallace D, et al. Altered irinotecan pharmacokinetics in pediatric high-grade glioma patients receiving enzyme-inducing anticonvulsant therapy. *Clin Cancer Res* 2002; 8: 2202-9
40. Murry DJ, Cherrick I, Salama V, et al. Influence of phenytoin on the disposition of irinotecan: a case report. *J Pediatr Hematol Oncol* 2002; 24: 130-3
41. Sanghani SP, Quinney SK, Fredenburg TB, et al. Hydrolysis of irinotecan and its oxidative metabolites, 7-ethyl-10-[4-N-(5-aminopentanoic acid)-carbonyloxy-camptothecin, by human carboxylesterases CES1A1, CES2, and a newly expressed carboxylesterase isoenzyme, CES3. *Drug Metab Dispos* 2004; 32: 505-11
42. Yamamoto W, Verweij J, de Buijn P, et al. Active transepithelial transport of irinotecan (CPT-11) and its metabolites by human intestinal Caco-2 cells. *Anticancer Drugs* 2001; 12: 419-32
43. Chester JD, Joel SP, Cheeseman SL, et al. Phase I and pharmacokinetic study of intravenous irinotecan plus oral ciclosporin in patients with fluorouracil-refractory metastatic colon cancer. *J Clin Oncol* 2003; 21: 1125-32

44. Takasuna K, Hagiwara T, Hirohashi M, et al. Inhibition of intestinal microflora beta-glucuronidase modifies the distribution of the active metabolite of the antitumor agent, irinotecan hydrochloride (CPT-11) in rats. *Cancer Chemother Pharmacol* 1998; 42: 280-6
45. Abigeres D, Chabot CG, Armand J-P, et al. Phase I and pharmacologic studies of the camptothecin analog irinotecan administered every 3 weeks in cancer patients. *J Clin Oncol* 1995; 13: 210-21
46. Alimonti A, Satta F, Pavese I, et al. Prevention of irinotecan plus 5-fluorouracil/leucovorin-induced diarrhoea by oral administration of neomycin plus bacitracin in first-line treatment of advanced colorectal cancer. *Ann Oncol* 2003; 14: 805-6
47. Lokiec F, Canal P, Gay C, et al. Pharmacokinetics of irinotecan and its metabolites in human blood, bile, and urine. *Cancer Chemother Pharmacol* 1995; 36: 79-82
48. Ratain MJ. Insights into the pharmacokinetics and pharmacodynamics of irinotecan. *Clin Cancer Res* 2000; 6: 3393-4
49. Santos A, Zanetta S, Cresteil T, et al. Metabolism of irinotecan (CPT-11) by CYP3A4 and CYP3A5 in humans. *Clin Cancer Res* 2000; 6: 2012-20
50. Mathijssen RH, Verweij J, de Bruijn P, et al. Effects of St. John's wort on irinotecan metabolism. *J Natl Cancer Inst* 2002; 94: 1247-9
51. Sai K, Kaniwa N, Ozawa S, et al. A new metabolite of irinotecan in which formation is mediated by human hepatic cytochrome P-450 3A4. *Drug Metab Dispos* 2001; 29: 1505-13
52. Slatter JG, Schaaf LJ, Sams JP, et al. Pharmacokinetics, metabolism, and excretion of irinotecan (CPT-11) following I.V. infusion of [(14)C]CPT-11 in cancer patients. *Drug Metab Dispos* 2000; 28: 423-33
53. Sparreboom A, de Jonge MJ, de Bruijn P, et al. Irinotecan (CPT-11) metabolism and disposition in cancer patients. *Clin Cancer Res* 1998; 4: 2747-54
54. Van Groenigen CJ, van der Vijgh WJ, Baars JJ, et al. Altered pharmacokinetics and metabolism of CPT-11 in liver dysfunction: a need for guidelines. *Clin Cancer Res* 2000; 6: 1342-6
55. Venook AP, Enders KC, Fleming G, et al. A phase I and pharmacokinetic study of irinotecan in patients with hepatic or renal dysfunction or with prior pelvic radiation: CALGB 9863. *Ann Oncol* 2003; 14: 1783-90
56. Ong SY, Clarke SJ, Bishop J, et al. Toxicity of irinotecan (CPT-11) and hepato-renal dysfunction. *Anticancer Drugs* 2001; 12: 619-25
57. Raymond E, Bojge V, Faivre S, et al. Dosage adjustment and pharmacokinetic profile of irinotecan in cancer patients with hepatic dysfunction. *J Clin Oncol* 2002; 20: 4303-12
58. Gandia D, Abigeres D, Armand JP, et al. CPT-11-induced cholinergic effects in cancer patients. *J Clin Oncol* 1993; 11: 196-7
59. Saliba F, Hagipantelli R, Misset JL, et al. Pathophysiology and therapy of irinotecan-induced delayed-onset diarrhea in patients with advanced colorectal cancer: a prospective assessment. *J Clin Oncol* 1998; 16: 2745-51
60. Dodds HM, Bishop JF, Rivory LP. More about: irinotecan-related cholinergic syndrome induced by coadministration of oxaliplatin. *J Natl Cancer Inst* 1999; 91: 91-2
61. Wasserman E, Cuvier C, Lokiec F, et al. Combination of oxaliplatin plus irinotecan in patients with gastrointestinal tumors: results of two independent phase I studies with pharmacokinetics. *J Clin Oncol* 1999; 17: 1751-9
62. Xu Y, Villalona-Calero MA. Irinotecan: mechanisms of tumor resistance and novel strategies for modulating its activity. *Ann Oncol* 2002; 13: 1841-51
63. Castellanos C, Aldaz A, Zufia L, et al. Biliary index accurately predict the severity of irinotecan (CPT-11) induced delayed diarrhea in colo-rectal cancer patients [abstract 648]. *Proc Am Soc Clin Oncol* 2003; 22: 162
64. Fittkau M, Voigt W, Holzhausen HJ, et al. Saccharic acid 1,4-lactone protects against CPT-11-induced mucosa damage in rats. *J Cancer Res Clin Oncol* 2004; 130: 388-94
65. Kehler DF, Sparreboom A, Verweij J, et al. Modulation of irinotecan-induced diarrhea by cotreatment with neomycin in cancer patients. *Clin Cancer Res* 2001; 7: 1136-41
66. Hennebell I, Terret C, Chatelut E, et al. Characterization of CPT-11 converting carboxylesterase activity in colon tumor and normal tissues: comparison with p-nitro-phenylacetate converting carboxylesterase activity. *Anticancer Drugs* 2000; 11: 465-70
67. Takeda Y, Kobayashi K, Akiyama Y, et al. Prevention of irinotecan (CPT-11)-induced diarrhea by oral alkalization combined with control of defecation in cancer patients. *Int J Cancer* 2001; 92: 269-75
68. Ikegami T, Ha L, Arimori K, et al. Intestinal alkalization as a possible preventive mechanism in irinotecan (CPT-11)-induced diarrhea. *Cancer Res* 2002; 62: 179-87
69. Prado D. A multinational comparison of racecadotril and loperamide in the treatment of acute watery diarrhoea in adults. *Scand J Gastroenterol* 2002; 37 (6): 656-62
70. Ukropec J, Pro B, Lozano R, et al. Refractory CPT-11 induced diarrhea in cancer patients: resolution with octreotide, four case studies [abstract]. *Proc Am Soc Clin Oncol* 2002; 21: 2904
71. Benson AB, Ajani JA, Catalano RB, et al. Recommended guidelines for the treatment of cancer treatment-induced diarrhea. *J Clin Oncol* 2004; 22: 2918-26
72. Barbounis V, Koumakis G, Vassilomanolakis M, et al. Control of irinotecan-induced diarrhea by octreotide after loperamide failure. *Support Care Cancer* 2001; 9: 258-60
73. Schwab M, Klotz U. Pharmacokinetic considerations in the treatment of inflammatory bowel disease. *Clin Pharmacokinet* 2001; 40: 723-51
74. Lenfers BH, Loeffler TM, Droegge CM, et al. Substantial activity of budesonide in patients with irinotecan (CPT-11) and 5-fluorouracil induced diarrhea and failure of loperamide treatment. *Ann Oncol* 1999; 10 (10): 1251-3
75. Karthaus M, Ballo H, Steinmetz T, et al. Budesonide for prevention of irinotecan (CPT-11)-induced diarrhea: results of a double-blind, placebo-controlled, multicenter, randomized phase-III-study in patients with advanced colorectal cancer (CRC) [abstract 2935]. *Proc Am Soc Clin Oncol* 2003; 22: 730
76. Sharma R, Tobin P. Management of chemotherapy-induced nausea, vomiting, oral mucositis, and diarrhoea. *Lancet Oncol* 2005; 9: 93-102
77. Michael M, Brittain M, Nagai J, et al. Phase II study of activated charcoal to prevent irinotecan-induced diarrhea. *J Clin Oncol* 2004; 22: 4410-7
78. Mansky PJ, Straus SE. St John's Wort: more implications for cancer patients. *J Natl Cancer Inst* 2002; 94: 1187-8
79. Mathijssen RH, Sparreboom A, Dumez H, et al. Altered irinotecan metabolism in a patient receiving phenytoin. *Anticancer Drugs* 2002; 13: 139-40
80. Kehler DF, Mathijssen RH, Verweij J, et al. Modulation of irinotecan metabolism by ketoconazole. *J Clin Oncol* 2002; 20: 3122-9
81. Minami H, Fujii H, Igarashi T, et al. Phase I and pharmacological study of a new camptothecin derivative, exatecan mesylate (DX-8951f), infused over 30 minutes every three weeks. *Clin Cancer Res* 2001; 7: 3056-64
82. Mitsui I, Kumazawa E, Hirota Y, et al. A new water-soluble camptothecin derivative, DX-8951f, exhibits potent antitumor activity against human tumors in vitro and in vivo. *Jpn J Cancer Res* 1995; 86: 776-82
83. Kajimura T. DX-8951f: single intravenous dose toxicity study in dogs. In house study report (8951 J-TOXO 19). Tokyo, Japan: Daiichi Pharmaceutical Co. Ltd; 1996

84. Braybrooke JP, Boven E, Bates NP, et al. Phase I and pharmacokinetic study of the topoisomerase I inhibitor, exatecan mesylate (DX-8951f), using a weekly 30-minute intravenous infusion, in patients with advanced solid malignancies. *Ann Oncol* 2003; 14: 913-21
85. Rowinsky EK, Johnson TR, Geyer CE, et al. DX-8951f, a hexacyclic camptothecin analog, on a daily-times-five schedule: a phase I and pharmacokinetic study in patients with advanced solid malignancies. *J Clin Oncol* 2000; 18: 3151-63
86. Jung LL, Ramanathan RK, Egorin MJ, et al. Pharmacokinetic studies of 9-nitrocamptothecin on intermittent and continuous schedules of administration in patients with solid tumors. *Cancer Chemother Pharmacol* 2004; 54: 487-96
87. Bailly C. Homocamptothecins: potent topoisomerase I inhibitors and promising anticancer drugs. *Crit Rev Oncol Hematol* 2003; 45: 91-108
88. de Jonge MJ, Verweij J, Loos WJ, et al. Clinical pharmacokinetics of encapsulated oral 9-aminocamptothecin in plasma and saliva. *Clin Pharmacol Ther* 1999; 65: 491-9
89. Ellerhorst JA, Bedikian AY, Smith TM, et al. Phase II trial of 9-nitrocamptothecin (RFS 2000) for patients with metastatic cutaneous or uveal melanoma. *Anticancer Drugs* 2002; 13: 169-72
90. Rowinsky EK, Rizzo J, Ochoa L, et al. A phase I and pharmacokinetic study of pegylated camptothecin as a 1-hour infusion every 3 weeks in patients with advanced solid malignancies. *J Clin Oncol* 2003; 21: 148-57
91. Saetern AM, Brandl M, Bakkelund WH, et al. Cytotoxic effect of different camptothecin formulations on human colon carcinoma in vitro. *Anticancer Drugs* 2004; 15: 899-906
92. Henwood JM, Brogden RN. Etoposide: a review of its pharmacodynamic and pharmacokinetic properties, and therapeutic potential in combination chemotherapy of cancer. *Drugs* 1990; 39: 438-90
93. Hande KR. Etoposide: four decades of development of a topoisomerase II inhibitor. *Eur J Cancer* 1998; 34: 1514-21
94. Hande KR. Topoisomerase II inhibitors. In: Giaccone G, Schilsky RL, Sordel P, editors. *Cancer chemotherapy and biological response modifiers*. Amsterdam: Elsevier, 2003: 103-25
95. Lipp H-P. Prevention and management of anticancer drug toxicity: the significance of clinical pharmacokinetics [thesis]. Jena: University of Jena, 1995
96. Hussain M, Vaishampayan U, Heilbrun LK, et al. A phase II study of rebeccamycin analog (NSC-655649) in metastatic renal cell cancer. *Invest New Drugs* 2003; 21: 465-71
97. Fortune JM, Osheroff N. Topoisomerase II as a target for anticancer drugs: when enzymes stop being nice. *Prog Nucleic Acid Res Mol Biol* 2000; 64: 221-53
98. Clark PI, Slevin ML. The clinical pharmacology of etoposide and teniposide. *Clin Pharmacokinet* 1987; 12: 223-52
99. Long BH. Mechanisms of action of teniposide (VM-26) and comparison with etoposide (VP-16). *Semin Oncol* 1992; 19: 3-19
100. Budman DR, Igwezie LN, Kaul S, et al. Phase I evaluation of a water-soluble etoposide prodrug, etoposide phosphate, given as a 5-minute infusion on days 1, 3, and 5 in patients with solid tumors. *J Clin Oncol* 1994; 12: 1902-9
101. Reif S, Kingreen D, Kloft C, et al. Bioequivalence investigation of high-dose etoposide and etoposide phosphate in lymphoma patients. *Cancer Chemother Pharmacol* 2001; 48: 134-40
102. Schacter LP, Igwezie LN, Seyedsadr M, et al. Clinical and pharmacokinetic overview of parenteral etoposide phosphate. *Cancer Chemother Pharmacol* 1994; 34 Suppl.: S58-63
103. Kaul S, Igwezie LN, Stewart DJ, et al. Pharmacokinetics and bioequivalence of etoposide following intravenous administration of etoposide phosphate and etoposide in patients with solid tumors. *J Clin Oncol* 1995; 13: 2835-41
104. Harvey VJ, Slevin ML, Joel SP, et al. The effect of dose on the bioavailability of oral etoposide. *Cancer Chemother Pharmacol* 1986; 16: 178-81
105. Hande KR, Krozely MG, Greco FA, et al. Bioavailability of low-dose oral etoposide. *J Clin Oncol* 1993; 11: 374-7
106. Aita P, Robieux I, Sorio R, et al. Pharmacokinetics of oral etoposide in patients with hepatocellular carcinoma. *Cancer Chemother Pharmacol* 1999; 43: 287-94
107. Millward MJ, Newell DR, Yuen K, et al. Pharmacokinetics and pharmacodynamics of prolonged oral etoposide in women with metastatic breast cancer. *Cancer Chemother Pharmacol* 1995; 37: 161-7
108. Chabot GG, Armand JP, Terret C, et al. Etoposide bioavailability after oral administration of the prodrug etoposide phosphate in cancer patients during a phase I study. *J Clin Oncol* 1996; 14: 2020-30
109. De Jong RS, Slijfer EAM, Uges DRA, et al. Conversion of the prodrug etoposide phosphate to etoposide in gastric juice and bile. *Br J Cancer* 1997; 76: 1480-3
110. Edick MJ, Gajjar A, Mahmoud HH, et al. Pharmacokinetics and pharmacodynamics of oral etoposide in children with relapsed or refractory acute lymphoblastic leukemia. *J Clin Oncol* 2003; 21: 1340-6
111. Toffoli G, Corona G, Basso B, et al. Pharmacokinetic optimisation of treatment with oral etoposide. *Clin Pharmacokinet* 2004; 43: 441-66
112. Joel SP, Shah R, Slevin ML. Etoposide dosage and pharmacodynamics. *Cancer Chemother Pharmacol* 1994; 34 Suppl.: S69-75
113. Joel S. The clinical pharmacology of etoposide: an update. *Cancer Treat Rev* 1996; 22: 179-221
114. Liu B, Earl HM, Poole CJ, et al. Etoposide protein binding in cancer patients. *Cancer Chemother Pharmacol* 1995; 36: 506-12
115. Nguyen L, Chatelut E, Chevreau C, et al. Population pharmacokinetics of total and unbound etoposide. *Cancer Chemother Pharmacol* 1998; 41: 125-32
116. D'Incalci M, Rossi C, Zucchetti M, et al. Pharmacokinetics of etoposide in patients with abnormal renal and hepatic function. *Cancer Res* 1986; 46: 2566-71
117. Stewart CF, Arbuck SG, Fleming RA, et al. Changes in the clearance of total and unbound etoposide in patients with liver dysfunction. *J Clin Oncol* 1990; 8: 1874-9
118. Schwinghammer TL, Fleming RA, Rosenfeld CS, et al. Disposition of total and unbound etoposide following high-dose therapy. *Cancer Chemother Pharmacol* 1993; 32: 273-8
119. Mross K, Bewermeier P, Kruger W, et al. Pharmacokinetics of undiluted or diluted high-dose etoposide with or without busulfan administered to patients with hematologic malignancies. *J Clin Oncol* 1994; 12: 1468-74
120. Kreis W, Budman DR, Vinciguerra V, et al. Pharmacokinetic evaluation of high-dose etoposide phosphate after a 2-hour infusion in patients with solid tumors. *Cancer Chemother Pharmacol* 1996; 38: 378-84
121. Zhuo X, Zheng N, Felix CA, et al. Kinetics and regulation of cytochrome P450-mediated etoposide metabolism. *Drug Metab Dispos* 2004; 32: 993-1000
122. Relling MV, Nemec J, Schuetz EG, et al. O-demethylation of epipodophyllotoxins is catalyzed by human cytochrome P450 3A4. *Mol Pharmacol* 1994; 45: 352-8
123. Hainsworth JD, Greco FA. Etoposide: twenty years later. *Ann Oncol* 1995; 6: 325-41
124. Hande KR, Wolff SN, Greco FA, et al. Etoposide kinetics in patients with obstructive jaundice. *J Clin Oncol* 1990; 8: 1101-7
125. Holthuis JJ, Van de Vyver FL, van Oort WJ, et al. Pharmacokinetic evaluation of increasing dosages of etoposide

- in a chronic hemodialysis patient. *Cancer Treat Rep* 1985; 69: 1279-82
126. Kiya K, Uozumi T, Ogasawara H, et al. Penetration of etoposide into human malignant brain tumors after intravenous and oral administration. *Cancer Chemother Pharmacol* 1992; 29: 339-42
 127. van der GA, Sonneveld P, Mans DR, et al. Intrathecal administration of etoposide in the treatment of malignant meningitis: feasibility and pharmacokinetic data. *Cancer Chemother Pharmacol* 1992; 29: 335-7
 128. Hande K, Messenger M, Wagner J, et al. Inter- and inpatient variability in etoposide kinetics with oral and intravenous drug administration. *Clin Cancer Res* 1999; 5: 2742-7
 129. Minami H, Ratain MJ, Ando Y, et al. Pharmacodynamic modeling of prolonged administration of etoposide. *Cancer Chemother Pharmacol* 1996; 39: 61-6
 130. Brooks DJ, Srinivas NR, Alberts DS, et al. Phase I and pharmacokinetic study of etoposide phosphate. *Anticancer Drugs* 1995; 6: 637-44
 131. Issell BF. The podophyllotoxin derivatives VP16-213 and VM26. *Cancer Chemother Pharmacol* 1982; 7 (2-3): 73-80
 132. Weiss RB. Hypersensitivity reactions. *Semin Oncol* 1992; 19: 458-77
 133. Alley E, Green R, Schuchter L. Cutaneous toxicities of cancer therapy. *Curr Opin Oncol* 2002; 14: 212-6
 134. Hudson MM, Weinstein HJ, Donaldson SS, et al. Acute hypersensitivity reactions to etoposide in a VEPA regimen for Hodgkin's disease. *J Clin Oncol* 1993; 11: 1080-4
 135. Ogle KM, Kennedy BJ. Hypersensitivity reactions to etoposide: a case report and review of the literature. *Am J Clin Oncol* 1988; 11: 663-5
 136. Jameson CH, Solanki DL. Stevens-Johnson syndrome associated with etoposide therapy. *Cancer Treat Rep* 1983; 67: 1050-1
 137. Hoetelmans RM, Schornagel JH, Bokkel Huinink WW, et al. Hypersensitivity reactions to etoposide. *Ann Pharmacother* 1996; 30: 367-71
 138. Nolte H, Carstensen H, Hertz H. VM-26 (teniposide)-induced hypersensitivity and degranulation of basophils in children. *Am J Pediatr Hematol Oncol* 1988; 10: 308-12
 139. Siderov J, Prasad P, De Boer R, et al. Safe administration of etoposide phosphate after hypersensitivity reaction to intravenous etoposide. *Br J Cancer* 2002; 86: 12-3
 140. Bernstein BJ, Troner MB. Successful rechallenge with etoposide phosphate after an acute hypersensitivity reaction to etoposide. *Pharmacotherapy* 1999; 19: 989-91
 141. Cersosimo RJ, Calarese P, Karp DD. Acute hypotensive reaction to etoposide with successful rechallenge: case report and review of the literature. *DIAP* 1989; 23: 876-7
 142. Wilson DB, Beck TM, Gundlach CA. Paclitaxel formulation as a cause of ethanol intoxication. *Ann Pharmacother* 1997; 31: 873-5
 143. Imrie KR, Couture F, Turner CC, et al. Peripheral neuropathy following high-dose etoposide and autologous bone marrow transplantation. *Bone Marrow Transplant* 1994; 13: 77-9
 144. McLeod HL, Baker DK, Pui CH, et al. Somnolence, hypotension, and metabolic acidosis following high-dose teniposide treatment in children with leukemia. *Cancer Chemother Pharmacol* 1991; 29: 150-4
 145. Nakanomyo H, Hiraoka M, Shiraya M. Mutagenicity tests of etoposide and teniposide [in Japanese]. *J Toxicol Sci* 1986; 11 Suppl. 1: 301-10
 146. Anderson RD, Berger NA. International Commission for Protection Against Environmental Mutagens and Carcinogens. Mutagenicity and carcinogenicity of topoisomerase-interactive agents. *Mutat Res* 1994; 309: 109-42
 147. Kollmannsberger C, Beyer J, Droz JP, et al. Secondary leukemia following high cumulative doses of etoposide in patients treated for advanced germ cell tumors. *J Clin Oncol* 1998; 16: 3386-91
 148. Duffner PK, Krischer JP, Horowitz ME, et al. Second malignancies in young children with primary brain tumors following treatment with prolonged postoperative chemotherapy and delayed irradiation: a Pediatric Oncology Group study. *Ann Neurol* 1998; 44: 313-6
 149. Horibe K, Matsushita T, Numata S, et al. Acute promyelocytic leukemia with t(15;17) abnormality after chemotherapy containing etoposide for Langerhans cell histiocytosis. *Cancer* 1993; 72: 3723-6
 150. Relling MV, Yanishevski Y, Nemec J, et al. Etoposide and antimetabolite pharmacology in patients who develop secondary acute myeloid leukemia. *Leukemia* 1998; 12: 346-52
 151. Zulian GB, Selby P, Milan S, et al. High dose melphalan, BCNU and etoposide with autologous bone marrow transplantation for Hodgkin's disease. *Br J Cancer* 1989; 59: 631-5
 152. Stine KC, Saylor RL, Sawyer JR, et al. Secondary acute myelogenous leukemia following safe exposure to etoposide. *J Clin Oncol* 1997; 15: 1583-6
 153. Houck W, Einhorn LH. Secondary leukemias in germ cell tumor patients undergoing autologous stem cell transplant utilizing high dose etoposide [abstract 1566]. *Proc Am Soc Clin Oncol* 2003; 22: 390
 154. Ratain MJ, Kaminer LS, Bitran JD, et al. Acute nonlymphocytic leukemia following etoposide and cisplatin combination chemotherapy for advanced non-small-cell carcinoma of the lung. *Blood* 1987; 70: 1412-7
 155. Pui CH, Relling MV. Topoisomerase II inhibitor-related acute myeloid leukaemia. *Br J Haematol* 2000; 109: 13-23
 156. Pui CH, Ribeiro RC, Hancock ML, et al. Acute myeloid leukemia in children treated with epipodophyllotoxins for acute lymphoblastic leukemia. *N Engl J Med* 1991; 325: 1682-7
 157. Mistry AR, Felix CA, Whitmarsh RJ, et al. DNA topoisomerase II in therapy-related acute promyelocytic leukemia. *N Engl J Med* 2005; 352: 1529-38
 158. Le Deley MC, Leblanc T, Shamsaldin A, et al. Risk of secondary leukemia after a solid tumor in childhood according to the dose of epipodophyllotoxins and anthracyclines: a case-control study by the Societe Francaise d'Oncologie Pediatric. *J Clin Oncol* 2003; 21: 1074-81
 159. Sandler ES, Friedman DJ, Mustafa MM, et al. Treatment of children with epipodophyllotoxin-induced secondary acute myeloid leukemia. *Cancer* 1997; 79: 1049-54
 160. Rodman JH, Abromowitch M, Sinkule JA, et al. Clinical pharmacodynamics of continuous infusion teniposide: systemic exposure as a determinant of response in a phase I trial. *J Clin Oncol* 1987; 5: 1007-14
 161. Lum BL, Kaubisch S, Yahanda AM, et al. Alteration of etoposide pharmacokinetics and pharmacodynamics by cyclosporine in a phase I trial to modulate multidrug resistance. *J Clin Oncol* 1992; 10: 1635-42
 162. Baker DK, Relling MV, Pui CH, et al. Increased teniposide clearance with concomitant anticonvulsant therapy. *J Clin Oncol* 1992; 10: 311-5
 163. Splinter TA, Holthuis JJ, Kok TC, et al. Absolute bioavailability and pharmacokinetics of oral teniposide. *Semin Oncol* 1992; 19: 28-34
 164. Bork E, Hansen M, Dombernowsky P, et al. Teniposide (VM-26), an overlooked highly active agent in small-cell lung cancer: results of a phase II trial in untreated patients. *J Clin Oncol* 1986; 4: 524-7
 165. Whitacre CM, Zborowska E, Gordon NH, et al. Topotecan increases topoisomerase IIalpha levels and sensitivity to treatment with etoposide in schedule-dependent process. *Cancer Res* 1997; 57: 1425-8

166. Bonner JA, Kozelsky TF. The significance of the sequence of administration of topotecan and etoposide. *Cancer Chemother Pharmacol* 1996; 39: 109-12
167. Dowlati A, Levitan N, Gordon NH, et al. Phase II and pharmacokinetic/pharmacodynamic trial of sequential topoisomerase I and II inhibition with topotecan and etoposide in advanced non-small-cell lung cancer. *Cancer Chemother Pharmacol* 2001; 47: 141-8
168. Hammond LA, Eckardt JR, Ganapathi R, et al. A phase I and translational study of sequential administration of the topoisomerase I and II inhibitors topotecan and etoposide. *Clin Cancer Res* 1998; 4: 1459-67
169. van Gijn R, Bokkel Huinink WW, Rodenhuis S, et al. Topoisomerase I/II inhibitor intoplicine administered as a 24h infusion: phase I and pharmacologic study. *Anticancer Drugs* 1999; 10: 17-23
170. Etievant C, Kruczynski A, Barret JM, et al. F 11782, a dual inhibitor of topoisomerases I and II with an original mechanism of action in vitro, and markedly superior in vivo antitumour activity, relative to three other dual topoisomerase inhibitors, intoplicin, aclarubicin and TAS-103. *Cancer Chemother Pharmacol* 2000; 46: 101-13
171. Caponigro F, Dittrich C, Sorensen JB, et al. Phase II study of XR 5000, an inhibitor of topoisomerases I and II, in advanced colorectal cancer. *Eur J Cancer* 2002; 38: 70-4

Correspondence and offprints: Dr Jörg T Hartmann, Department of Oncology/Hematology/Immunology/Pneumology/Rheumatology, UKT-Medical Center II, Eberhard-Karls-University Tübingen, Otfried-Mueller-Str. 10, 72076 Tübingen, Germany.
E-mail: joerg.hartmann@med.uni-tuebingen.de